Problem Space

Fossil fuels

- Dominant energy source today. They accounted for 86% of world's energy mix in 2024.
- High greenhouse gas emissions drive climate change. In 2024, atmospheric CO₂ from burning fossil fuels reached 423 ppm, with emissions of 37.4 billion tons.
- Non-renewable, cause environmental pollution, and threats to biodiversity.

The Future of Energy: Green Power, Clean Future

Microalgae as a source of biofuel

- ✓ Unicellular organisms: Can be freshwater or marine and are fast growing.
- ✓ Produce oil or biolipids that can be converted to Biofuel.
- ✓ Renewable and carbon neutral absorbs CO₂ as they grow
- ✓ Sustainable: Rapid growth, 20x more oil per acre than crops
- ✓ Water efficient: Grows on non-arable land and wastewater no food competition

Transesterification Reaction to Convert Biolipids to Biodiese H-C-OH + H3C 0 Image created by Finalist using Canva, 2025

Current Challenges

Operational difficulties:

- > Low and inconsistent biomass yield
- > Low lipid content: difficult to maintain growth for high lipid production
- > Harvesting and extraction difficulties
- > Competition with cheaper fossil fuels

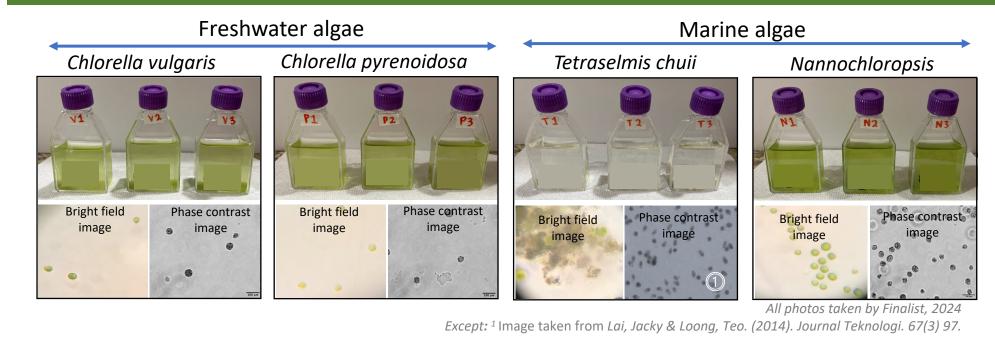
Engineering Goals

- o Improve Biomass Yield: Vitamin B1 and B Complex may help increase algal metabolism by acting as cofactors in the metabolic pathways. Vitamin C may increase photosynthesis and protects the cells from being damaged. Indole 3-acetic acid (IAA) is a plant hormone that could help cell division.
- Improve Oil Production: Salt stress may increase the amount of lipids produced per cell, by changing the metabolic pathway from converting CO₂ from air to lipids instead of sugars and proteins.

Questions Asked

- A. Which freshwater and marine microalgae produce the most biomass in their standard growth conditions?
- B. Will the addition of Vitamins B, C and plant hormone to standard growth media increase biomass?
- C. Will saline stress given to freshwater microalgae increase their lipid production?

Materials



4. Other materials:

Algal growth media – Fresh

water (Bold's basal media), and

saltwater media (Guillard and

500 ml wide mouth bottles

DPBS (buffer for rinsing cells)

Ryther's f/2 media)

25 ml T-Flasks

Secchi Sticks

10 ml Syringe

3 ml dropper

15 mL test tubes

• 100-1000 µl pipette

Disposable pipette tips

Microcentrifuge tubes

Countess cell counter

Fluorescence microscope

Fluorescence plate reader

• 20-200 μl pipette

2-20 µl pipette

- 1. Microalgae:
- · Chlorella vulgaris- culture and as sodium alginate algal beads
- · Chlorella pyrenoidosa
- Nannochloropsis
- · Tetraselmis chuii
- 2. Additives:
- Vitamin B1 (6 mg/mL) Vitamin B Complex
- Contains five key B-vitamins including B12 (1.2 mg/mL)
- Vitamin C (22.5 mg/mL)
- Indole 3-acetic acid (IAA, a plant growth hormone)
- (0.1mg/100mg solid) Sea Salt
- 3. Chemicals:
- Acetone Nile red dye

Micro'fat'ories: Increasing Microalgal Biomass and Lipids for Biofuel Production

Experimental Methods

Experiment 1 Comparing growth kinetics of different microalgae Freshwater Saltwateı media + media + microalgae Aeration and sunlight

Experiment 2

Effect of individual additives on photosynthesis by Chlorella

vulgaris

Experiment 3

Effect of vitamin B complex, C and IAA on the fastest

growing freshwater and marine algae from Experiment 1

Experiments 4 and 5

Effect of vitamin B complex, C and sea salt on growth

and lipid production by microalgae when added

either simultaneously (Experiment 4) or sequentially

(Experiment 5)

days

Aeration and sunlight

Compare Growth for 10 Days

Experiment 4

Experiment 5

Vitamin B complex + C

Vitamin

B complex + C

Density Measurement

algae = consumes CO2 + raises pH

_ Vit B complex + Vit C + IAA

Image created by Finalist using Canva, 2025

Aeration and sunlight

Compare Growth for 10 Days

Image created by Finalist using Canva, 2025

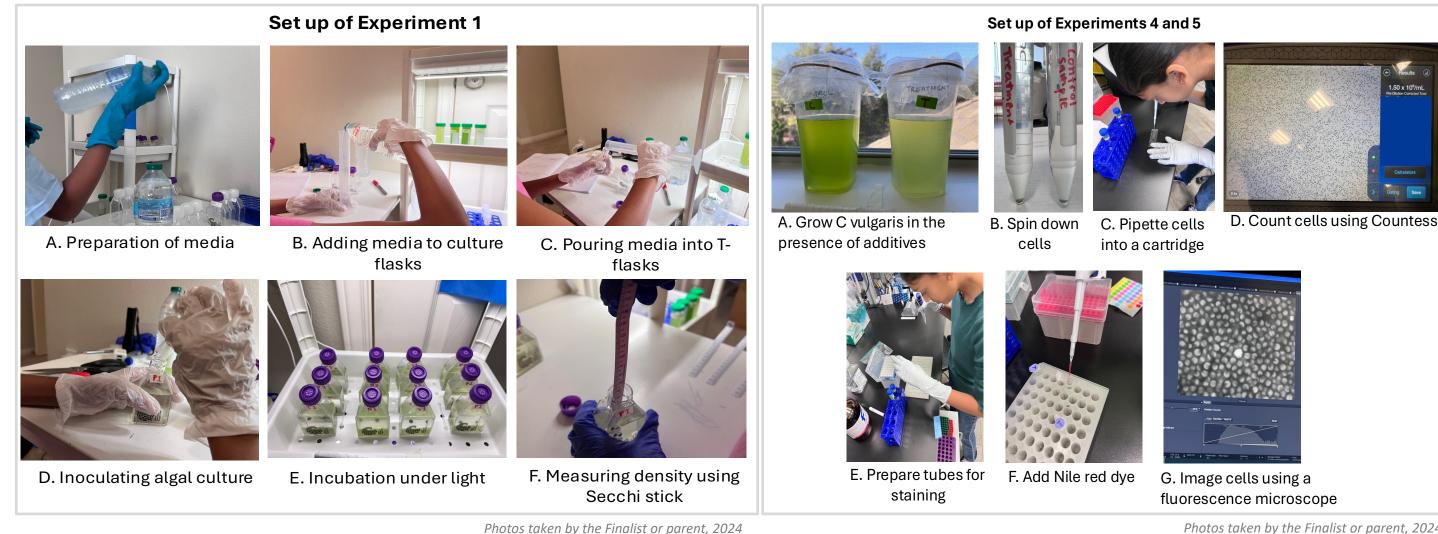
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Image created by Finalist using Canva, 2025

Quantitation of

oil production

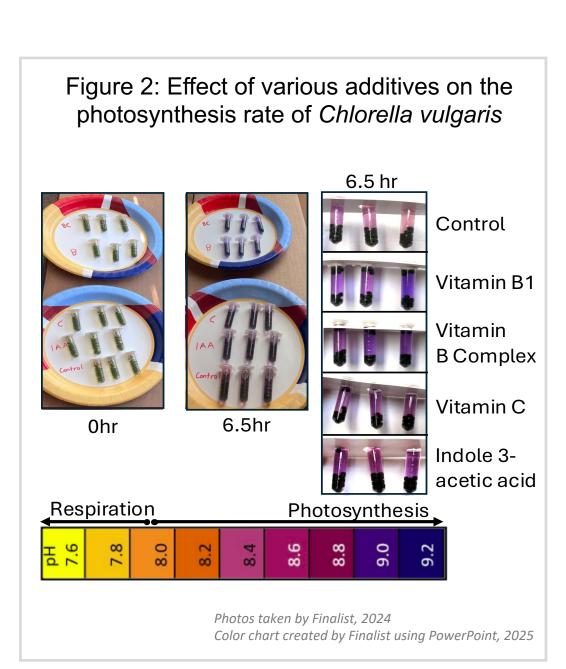
Set up of Experiment



Experimental Setup

Data

Figure 1: Comparing growth kinetics of the microalgae: Chlorella vulgaris, Chlorella pyrenoidosa, Nannochloropsis, and Tetraselmis → C.pyrenoidosa -- Nannochloropsi Day 0 Day 1 Day 2 Day 5 Day 5 Day 6 Day 7 Day 9 Day 10 Day 12 Day 13 Day 15 Graph created by Finalist using Excel, 2025



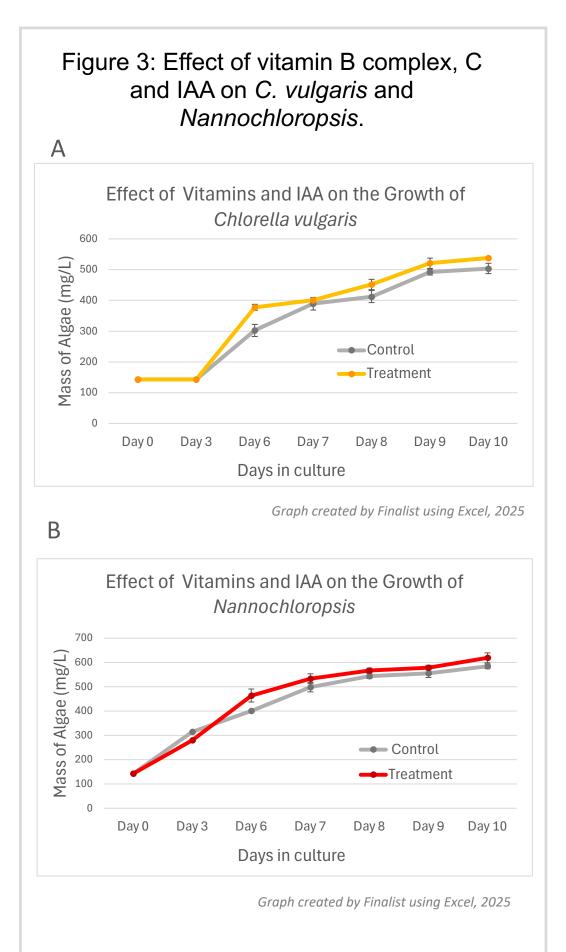


Figure 4a: Effect of Vitamin B complex, C, and the addition of salt simultaneously on the growth of Chlorella vulgaris

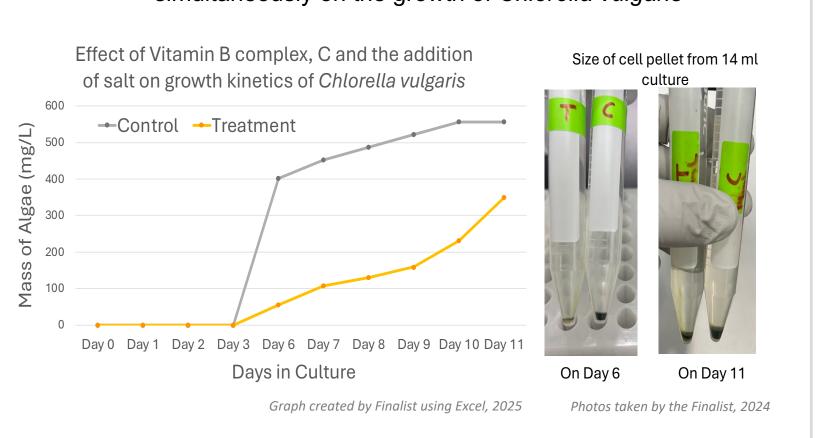


Figure 4b: Effect of Vitamin B complex, C, and the addition of salt simultaneously on lipid production by Chlorella vulgaris

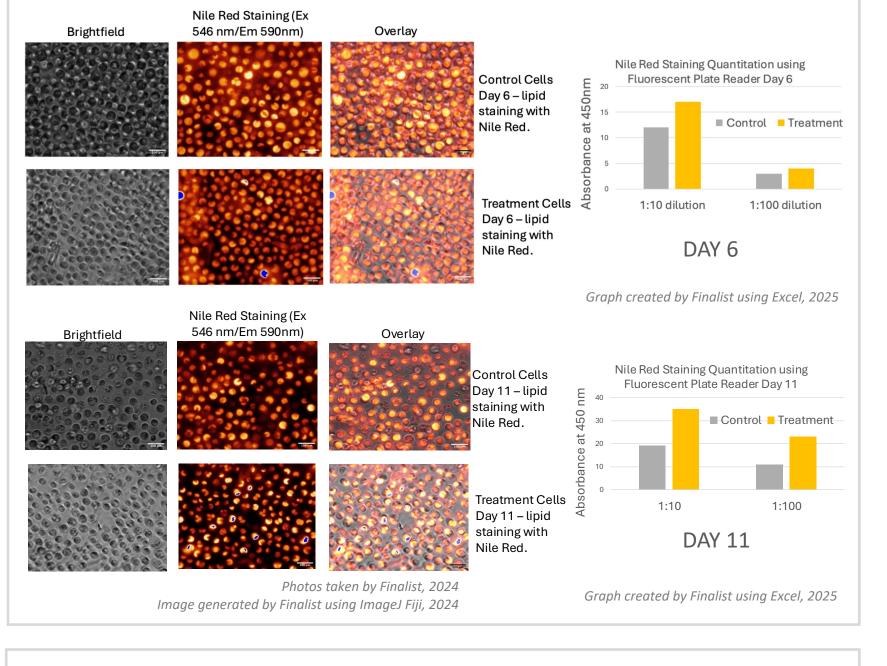
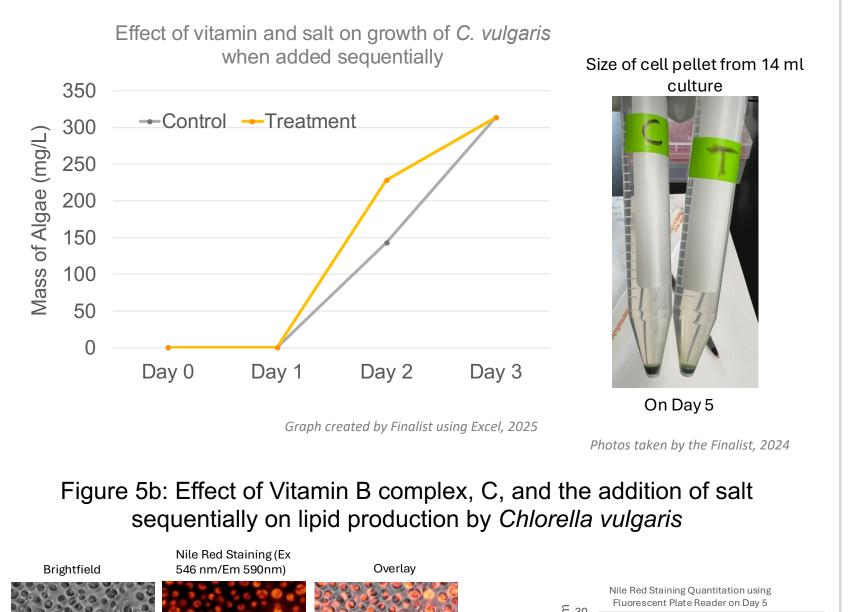


Figure 5a: Effect of Vitamin B complex, C, and the addition of salt sequentially on the growth of Chlorella vulgaris



Day 5 – lipid staining with 1:20 dilution

DAY 5

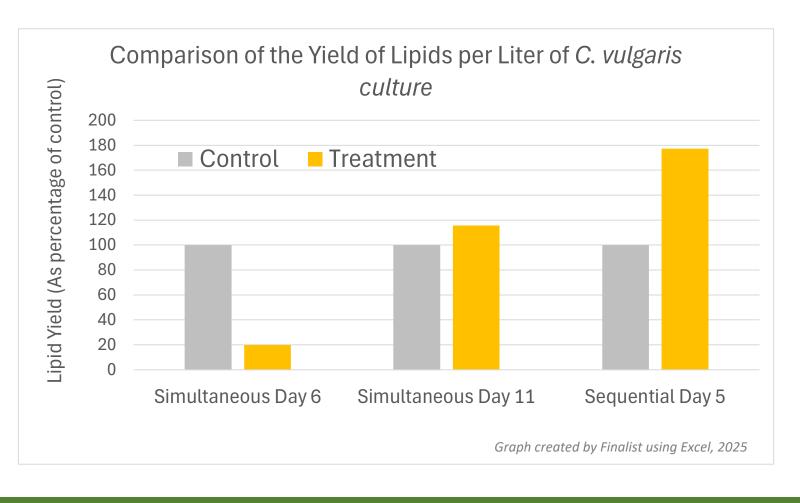
Graph created by Finalist using Excel, 2025 Photos taken by Finalist, 2024 Image generated by Finalist using ImageJ Fiji, 2024

Results

- Results from this study showed that Chlorella vulgaris and Nannochloropsis have high growth rates.
- The addition of vitamins B complex and C increased the rate of photosynthesis, and co-treatment of vitamins and plant hormone IAA increased biomass yield of Nannochloropsis and Chlorella vulgaris by 6% and 7%, respectively.
- Treatment with salt and vitamins simultaneously lowered biomass yield, however, increased lipid production by ~84% per mg of microalgae by Day 11.
- Treatment with vitamins for first three days followed by salt stress for next two days did not lower the biomass yield, however, it increased production of lipids by 77% per mg of microalgae by Day 5.

Conclusion

- Based on the results, it can be concluded that vitamins and high salt stress help increase algal growth and lipid production in a positive way.
- If vitamins and salt are added simultaneously, even though there is an increase in lipid production per cell, the amount of biomass will be less, resulting in low total lipid yield in the
- The better way would be a two-step process in which the addition of vitamins come first for three days, followed by the addition of salt. This would result in the highest total lipid yield in the culture.



Future Direction

- Scaling up the 2-step process to a manufacturing scale.
- Optimization of Step 1 (vitamin treatment) and Step 2 (salt stress) to be done based on the typical duration for each stage in the microalgal life cycle in a bioreactor:
 - Lag phase (adaptation): about 1 day
- Exponential growth phase (vitamin treatment): 4 to 7 days depending on species and conditions.
- Stationary phase (addition of salt): around day 6 to 8.
- Harvesting process: Can vary from 1 hour to a day.
- Cleaning/sterilization and preparation for the next cycle: Can take up to multiple days depending on size of the bioreactor
- Test if low salt stress increases oil production by marine microalgae Nannochloropsis.

Scaling Up for Production

